STUDIES ON BIOLOGICALLY IMPORTANT COPPER(II) / MANGANESE(II) / URANYL(II) – NORVALINE BINARY COMPLEXES

Brij Bhushan Tewari

Department of Chemistry, Faculty of Natural Sciences, University of Guyana, P. O. Box: 101110, Georgetown, Guyana
brijtew@yahoo.com

In coordination compounds studies, a knowledge of the magnitude of the stability constants of complexes is necessary for preliminary quantitative treatment. Described herein is a method that involves the use of advanced ionophoretic technique for the study of the equilibria in binary complex systems in solution. This method is based upon the migration of a spot of the metal ion on a paper strip at different pH values of background electrolyte containing 0.1 M perchloric acid and 0.01 M norvaline. A graph of pH against mobility provides information about the nature of the complexation and helps in calculating stability constants. Using this method, the stability constants of binary complexes metal(II) – norvaline have been determined to be (8.11 ± 0.02, 7.03 ± 0.09); (3.77 ± 0.11, 2.39 ± 0.07) and (7.59 ± 0.05, 6.17 ± 0.04) (log K values) for Cu(II), Mn(II) and UO₂(II) complexes, respectively, at 35ºC.

Key words: electrophoretic technique; complexation reactions; copper(II) complexes; manganese(II) complexes; uranyl(II) complexes; norvaline; stability constants

1. INTRODUCTION

Data on the complexation of essential metal ions and the bioactive ligands norvaline gives an important insight into many physiochemical processes. Norvaline is an amino acid (C₅H₁₁NO₂) isomeric with valine, and is commonly made synthetically. It has several significances in biological systems (1–9). The importance of metal ions to vital functions in living systems and for the well being of living organisms is now well established. Copper and manganese are classified as essential metals, while uranyl is a toxic metal. Average copper and manganese content of a healthy human (body weight, 70 kg) are 0.100 and 0.012 g/individual, respectively. The normal recommended amount of uranium in human diet is 0.001 – 0.002 mg/day (10). Copper(II), manganese(II) and uranyl(II) have significant biomedical applications, but are toxic at higher concentration (11–27).
The shortcomings of paper ionophoretic technique like variation in the temperature during electrophoresis, capillary flow on paper, electro-osmosis, and adsorption effects on the mobility of charged moieties, are well known (28). In this work we describe a technique that overcomes these limitations. The technique is very convenient, simple in use, and yields results in fair agreement with the accepted literature values.

Recent publications from our laboratory (29–31) described a new method for the study of binary complexes by using paper ionophoretic technique. The present work is an extension of the technique, and reports an observation on the determination of the stability constant values of binary complexes of copper(II)/manganese(II)/uranyl(II) – norvaline.

2. EXPERIMENTAL SECTION

2.1. Apparatus

A Systronic (Naroda, India), Model 604, electrophoresis systems was used. The apparatus consisted of a PVC moulded double tanks vessel. A significant change in the instrument were made: two hollow rectangular plates covered with thin polythene sheets were used through which thermostated water was run for controlling the temperature, and the tanks were closed with a transparent PVC moulded lid. The whole assembly was tight, which prevented moisture changes, which may upset the equilibria in a paper strips. The assembly design thus kept to a minimum the disturbing effect of evaporation from the unwanted liquid flow in the paper. Each electrolyte tank contained a separate electrode chamber in which the anode and cathode were placed, respectively.

Whatman No. 1 filter papers for chromatography were used for the purpose of electrophoresis. Elico (Hyderabad, India) Model L1–10 pH meter (glass and calomel electrode assembly, working on 220 V/50 Hz established a.c. mains) was used for pH measurements. The pH meter was calibrated with buffer solution of pH 7.0.

2.2. Chemicals

Solutions of copper(II), manganese(II) and uranyl(II) metal perchlorate were prepared by preliminary precipitation of metal carbonates from 0.1 mol L\(^{-1}\) solution of sodium carbonate (AnalaR grade; BDH, Poole, UK). The precipitate was thoroughly washed with boiling water and treated with predetermined amounts of 1% perchloric acid. The resulting mixture was heated to boiling on a water bath and was then filtered. The metal contents of the filtrates were determined and the final concentration was kept at 5.0×10\(^{-3}\) mol L\(^{-1}\).

Metal spots were detected on the paper using a 0.1% solution of 1 – (2-pyridylazo) – 2 – naphthol (PAN) (E. Merck, Darmstadt, Germany). A saturated aqueous solution (0.9 ml) of silver nitrate was diluted with acetone to 20 ml. Glucose, as the black spot, was detected by spraying with this solution and then with 2% ethanolic solution of sodium hydroxide.

2.3. Background electrolyte

The background electrolyte used in the study of binary complexes were 0.1 mol L\(^{-1}\) perchloric acid and 0.01 mol L\(^{-1}\) norvaline. Stock solution of 5.0 mol L\(^{-1}\) perchloric acid was prepared from 70% solution (SDS, AnalaR grade). 2.0 mol L\(^{-1}\) sodium hydroxide (AnalaR grade) and 0.5 mol L\(^{-1}\) norvaline (BDH, Poole, UK) solutions were prepared. Each solution was standardized using appropriate methods.

2.4. Procedure

The hollow base plate in the instrument was verified to be horizontal with spirit level. A 150 mL volume of background electrolyte was placed in each tank of the electrophoretic apparatus. The levels of the two tank solutions were equalized by a siphon. These precaution were taken to stop any gravitational and hydrodynamic flow. Paper strips (Whatman No. 1) of 30×1 cm\(^2\) size were soaked in background electrolyte and the excess electrolyte solution was blotted by pressing them gently within the folds of dry filter paper sheets. Paper strips in duplicate were spotted with metal ions and glucose in the centre with a micropipette and were subsequently placed on the base plate and sandwiched under the upper hollow metallic plate with the ends of the strips immersed in the tank solutions on both sides. A potential difference of 220 V was applied between the tank solutions, and electrophoresis was carried out for 60 minutes. Afterward the strips were removed with the help of glass rod, dried, and the spots were developed. The distance recorded in duplicate strips differed ± 5% and the average was calculated. The distances
travelled towards anode were given a negative sign and those towards the cathode positive. The actual distance of the sample spot moved was corrected for the distance travelled by the reference glucose spot. The mobilities were calculated by dividing the movement with the potential gradient and expressed in cm² V⁻¹ min⁻¹.

RESULTS

The ionophoretic mobility of metal spot against pH yields a curve with a number of plateaus, as shown in Figure 1. Every plateau indicates the formation of a certain complex species. A plateau is an indication of a pH range where speed is practically constant. The first one in the beginning corresponds to a region in which metal ions are uncomplexed. In this region of low pH, the concentration of $[\text{CH}_3 \, \text{CH}_2 \, \text{CH}_2 \, \text{CH} \,(\text{NH}_3^+) \, \text{COOH}]$ species of norvaline is maximum and this species is non-complexing. Beyond this range, metal ion spots have progressively decreasing mobility, and complexation of metal ions should be taking place with anionic species of norvaline whose concentration increases progressively with the increase of pH. Figure 1 shows three plateaus for copper(II), manganese(II) and uranyl(II) metal ions. All three metal ions form two complexes with norvaline anion. The literature also assigns prominent liganding property to the zwitterion species [32]. It is therefore assumed that the anionic species of norvaline $[\text{CH}_3 \, \text{CH}_2 \, \text{CH}_2 \, \text{CH} \,(\text{NH}_2) \, \text{COO}^-]$ has complexed with the metal ions to form different complex species.

Figure 1 reveals that $\text{Cu}^{2+}$, $\text{Mn}^{2+}$ and $\text{UO}_2^{2+}$ metal ions form their first complex movement towards negative electrode. Hence one $[\text{CH}_3 \, \text{CH}_2 \, \text{CH}_2 \, \text{CH}_,(\text{NH}_2) \, \text{COO}^-]$ must have combined with $\text{Cu}^{2+}$, $\text{Mn}^{2+}$ and $\text{UO}_2^{2+}$ metal ions to give $[\text{Cu} \, \{\text{CH}_3 \, \text{CH}_2 \, \text{CH}_2 \, \text{CH} \,(\text{NH}_2) \, \text{COO}\}]^+$, $[\text{Mn} \, \{\text{CH}_3 \, \text{CH}_2 \, \text{CH}_2 \, \text{CH} \,(\text{NH}_2) \, \text{COO}\}]^+$ and $[\text{UO}_2 \, \{\text{CH}_3 \, \text{CH}_2 \, \text{CH}_2 \, \text{CH} \,(\text{NH}_2) \, \text{COO}\}]^+$ complex cations, respectively. With further increase of pH, the mobility of all three metal ions decreased, giving rise to a third plateau with zero mobility, indicating its neutral nature. The third plateau in each case is due to a 1:2 metal – ligand complex. Hence, two $[\text{CH}_3 \, \text{CH}_2 \, \text{CH}_2 \, \text{CH}_,(\text{NH}_2) \, \text{COO}^-]$ must have combined with $\text{Cu}^{2+}$, $\text{Mn}^{2+}$ and $\text{UO}_2^{2+}$ to give $[\text{Cu} \, \{\text{CH}_3 \, \text{CH}_2 \, \text{CH}_2 \, \text{CH} \,(\text{NH}_2) \, \text{COO}\}]_2^+$, $[\text{Mn} \, \{\text{CH}_3 \, \text{CH}_2 \, \text{CH}_2 \, \text{CH} \,(\text{NH}_2) \, \text{COO}\}]_2^+$ and $[\text{UO}_2 \, \{\text{CH}_3 \, \text{CH}_2 \, \text{CH}_2 \, \text{CH} \,(\text{NH}_2) \, \text{COO}\}]_2^+$ metal complexes. In view of the above observation, the complexation of metal ions with the norvaline anion may be given as:

$$M^{2+} + L^- \overset{K_1}{\longleftrightarrow} ML^+, \quad (1)$$

$$ML^+ + L^- \overset{K_2}{\longleftrightarrow} ML_2, \quad (2)$$

wherein $M^{2+}$ represents the $\text{Cu}^{2+}$, $\text{Mn}^{2+}$ and $\text{UO}_2^{2+}$ metal cations, $[L^-]$ is the norvaline anion; and $K_1$ and $K_2$ are the first and second stability constants, respectively.

Fig. 1. Mobility curve for the metal(II)–norvaline systems.

- $\text{Cu}(\text{II})$–norvaline,
- $\text{Mn}(\text{II})$–norvaline,
- $\text{UO}_2$(II)–norvaline. Concentration of Cu(II), Mn(II) and UO2(II) = 0.005 mol l⁻¹. The paper strips were spotted with 0.1 µl of sample solutions and glucose (for making osmotic corrections)

The metal spot on the paper is thus a combination of the uncomplexed metal ions; 1:1 complex, and 1:2 complex. The spot is moving under the influence of the electric field, and the overall mobility is given by the equation of Jokl [33]:

$$U = \frac{\sum u_{xp} \cdot \beta_{xp} [HpL]^x}{\sum \beta_{xp} [HpL]^x}, \quad (3)$$

wherein $[HpL]^x$ is the concentration of general complex species; $\beta_{xp}$ is the overall mobility constant of the complex; $U_{xp}$ is the speed of the general complex $[M(HpL)]^x$ present in the combination. On taking into consideration different equilibria, the above equation is transformed into the following form:

\[ M_{\text{Cu}^{2+}}^{\text{Cu}^{2+}} + L^- \overset{K_1}{\longleftrightarrow} ML^+, \quad (1) \\
ML^+ + L^- \overset{K_2}{\longleftrightarrow} ML_2, \quad (2) \]

\[ M_{\text{Cu}^{2+}}^{\text{Cu}^{2+}} + L^- \overset{K_1}{\longleftrightarrow} ML^+, \quad (1) \\
ML^+ + L^- \overset{K_2}{\longleftrightarrow} ML_2, \quad (2) \]
wherein $u_0$, $u_1$, and $u_2$ are the mobilities of the uncomplexed metal ions, 1:1 and 1:2 metal complexes, respectively. Eq. (4) was used for the determination of the stability constants of the metal ions with norvaline. For calculating the first stability constant, $K_1$, the region between first and second plateau is pertinent. The overall mobility $U$ is equal to the arithmetic mean of the mobility of the uncomplexed metal ion, $u_0$, and that of the first complex, $u_1$, at a pH, where

$$K_1 = 1/[\text{CH}_3\text{CH}_2\text{CH}_2\text{CH}-(\text{NH}_2)\text{COO}^-].$$

With the help of dissociation constants of norvaline [$k_1 = 10^{2.31}$, $k_2 = 10^{9.65}$ (electrophoretically obtained values)], the concentration of the norvaline anion $[L^-]$ is determined for the pH using the following equation, from which $K_1$, can be calculated.

$$[L^-] = \frac{[L_T]}{1 + [H]/k_2 + [H]^2/k_1 \cdot k_2},$$

wherein $[L_T]$ is the total concentration of norvaline (0.01 mol L$^{-1}$); and $k_1$ and $k_2$ are the first and second dissociation constants of pure norvaline, respectively. The dissociation of norvaline can be represented as:

$$[\text{CH}_3\text{CH}_2\text{CH}_2\text{CH}-(\text{NH}_3^+)\text{COOH}]$$
$$- H^+ \uparrow k_1$$

$$[\text{CH}_3\text{CH}_2\text{CH}_2\text{CH}-(\text{NH}_3^+)\text{COO}^-]$$
$$- H^+ \uparrow k_2$$

$$[\text{CH}_3\text{CH}_2\text{CH}_2\text{CH}-(\text{NH}_2)\text{COO}^-]$$

The stability constant of second complex, $K_2$, can be calculated by taking into consideration the region between the second and third plateau of the mobility curve. These calculated values are given in the Table 1.

### Table 1

<table>
<thead>
<tr>
<th>Metal ions</th>
<th>Stability constants</th>
<th>Complexes</th>
<th>$pK_1$ and $pK_2$</th>
<th>$\Delta pK$</th>
<th>Literature</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cu$^{2+}$</td>
<td>$K_1$</td>
<td>ML$^+$</td>
<td>8.11 ± 0.02</td>
<td>8.17 (34)</td>
<td></td>
</tr>
<tr>
<td></td>
<td>$K_2$</td>
<td>ML$^2$</td>
<td>7.03 ± 0.09</td>
<td>6.87 (34)</td>
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</tr>
<tr>
<td>Mn$^{2+}$</td>
<td>$K_1$</td>
<td>ML$^+$</td>
<td>3.77 ± 0.11</td>
<td>3.30 (35)</td>
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</tr>
<tr>
<td></td>
<td>$K_2$</td>
<td>ML$^2$</td>
<td>2.39 ± 0.07</td>
<td>1.89 (35)</td>
<td></td>
</tr>
<tr>
<td>UO$_2$$^{2+}$</td>
<td>$K_1$</td>
<td>ML$^+$</td>
<td>7.59 ± 0.05</td>
<td>7.59 ± 0.05</td>
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<tr>
<td></td>
<td>$K_2$</td>
<td>ML$^2$</td>
<td>6.17 ± 0.04</td>
<td>6.17 ± 0.04</td>
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</tr>
</tbody>
</table>

Ionic strength = 0.1 mol l$^{-1}$; temperature = 35ºC; norvaline anion: [CH$_3$CH$_2$CH$_2$CH(NH$_2$)COO$^-$]; M = metal cations (Cu$^{2+}$, Mn$^{2+}$ and UO$_2$$^{2+}$); L = ligand (norvaline); conc. of metal cations = 0.005 mol l$^{-1}$; pH of the solution was maintained by addition of sodium hydroxide. The paper strips were spotted with 0.1 µl of sample solution and glucose (for making osmotic correction).

It is clear from the Table 1 that the calculated stability constants are similar to those reported in the literature. The slight deviation in the values obtained from different sources is mainly due to the differences in temperature and ionic strength used by different workers. The stability constants of metal complexes can be very easily calculated by this technique, therefore the present method is advantageous over other methods (viz, polarographic, potentiometric, solubility etc.). It can also be concluded that norvaline may be used to reduce the level of copper(II), manganese(II) and uranyl(II) in biological systems. This simple electrophoretic technique has thus proved to be helpful in deciding whether a mixed complex system is formed, and if it is formed its stability constants can readily be calculated.
REFERENCES


